



Soil Carbon Storage in Reforested Tropical Pastures

Erika Marín-Spiotta

GREF Fellow

Ecosystem Sciences Division

Dept. of Environmental Science, Policy & Management

U.C. Berkeley

View From Below: Soils can be sexy, too (and why you should care)



- Need a better understanding of factors controlling soil C dynamics and its effect on atmospheric CO₂
- Scientific and political interest in C sequestration in soils
- Rehabilitation degraded soils: agricultural and natural
- SOM as fertility source
- Bioremediation

Land Use Change: Large-Scale Field Experiments

LUC affects factors controlling soil C storage:

Physical: occlusion, barriers to decomposers; moisture, temperature, O₂

Chemical: litter quality, sorption,

Biological: decomposer community

Interactions of all of the above to slow down organic matter decomposition.



Reforested Tropical Pastures

- Differences in quantity and quality of above and belowground litter inputs.
- Differences in $^{13}\text{C} / ^{12}\text{C}$ of tropical forage grasses (**C₄ photosynthesis**) and woody vegetation (**C₃**).

C₄ plants
(average $\delta^{13}\text{C} = -12\text{‰}$)



C₃ plants
(average $\delta^{13}\text{C} = -25\text{‰}$)



Secondary forests dominate tropical (and temperate!) landscape.

- Deforestation main land use studied in tropics
- Puerto Rico is at opposite end of land conversion: reforestation important process
- Reforestation important ecologically and economically



Research Objectives:

- To examine mechanisms that lead to soil C storage in tropical soils
- To describe general pattern in soil C accumulation or loss with reforestation of tropical pastures



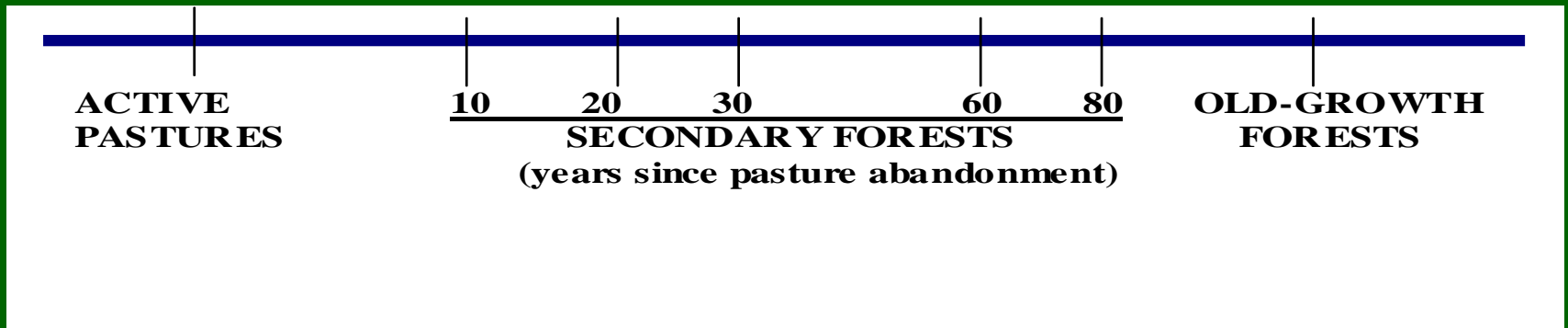
PUERTO RICO



0 20 km
0 12 mi



CHRONOSEQUENCE APPROACH



- Wet subtropical forest (400-600 masl).
- 7 age classes, 3 site replicates per age for a total of 21 sites



Field Sampling Summers 2001, 2002, 2003

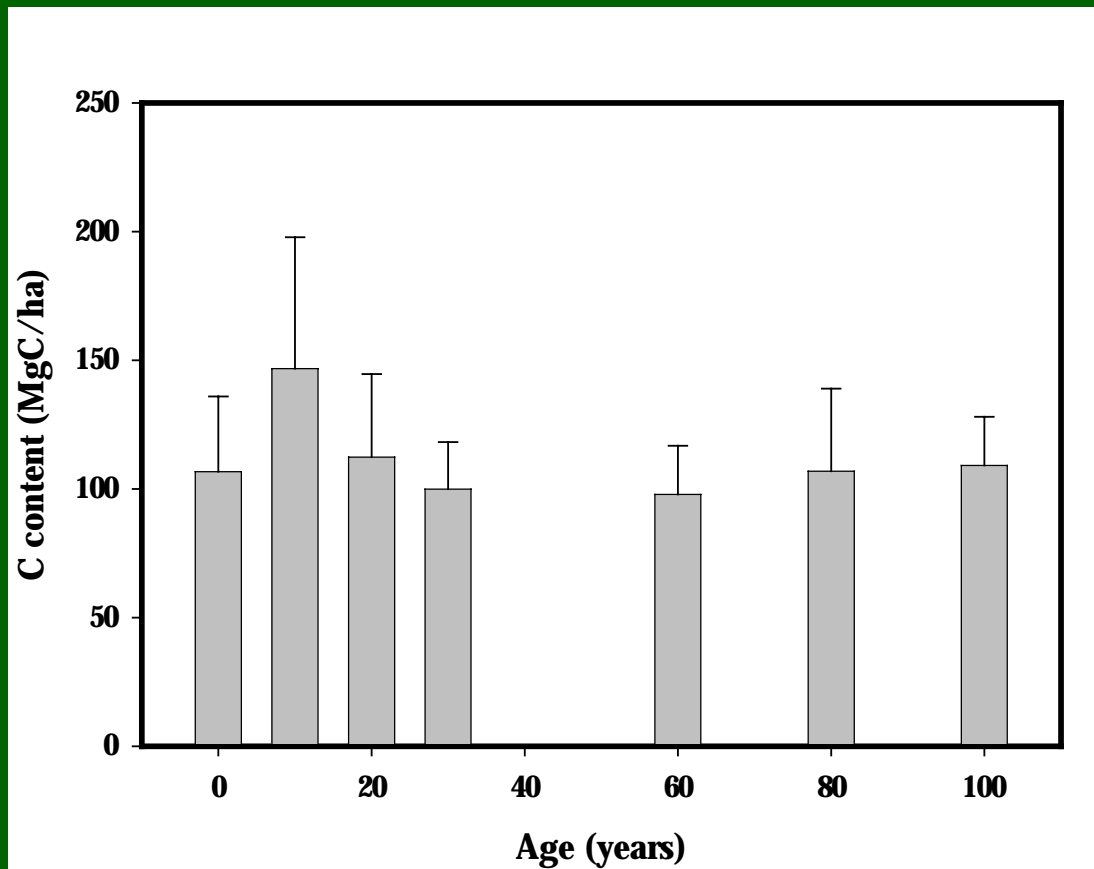
- Collect soils every 10 cm to a 1 m depth, 3 soil pits per site
- Collect roots, forest floor
- Litterfall (biweekly)
- Tree basal area measurements and species identification





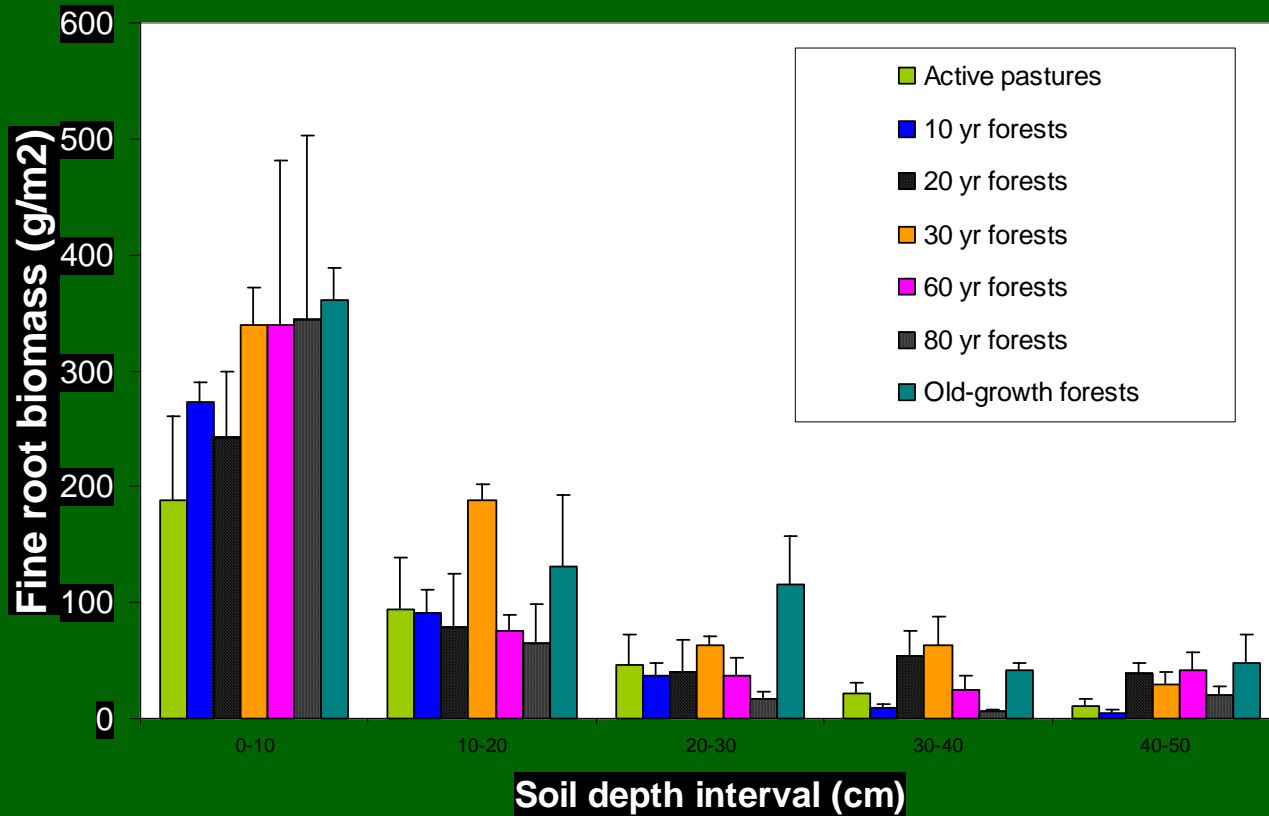
- **Objective 1 : Changes in soil C with forest succession**
- **Deforestation usually results in initial loss of soil organic matter (C and nutrients)**
- **Reforestation and afforestation as policy actions for increasing C sequestration in tree biomass and soils**
- **Reforestation: both net changes in soil C stocks and no changes have been measured**
- **Land use type (agriculture vs. pasture), intensity and duration matter**

Age or land use did not have a significant effect on total C content in the top 1 m of soil (or top 30 cm- data not shown).

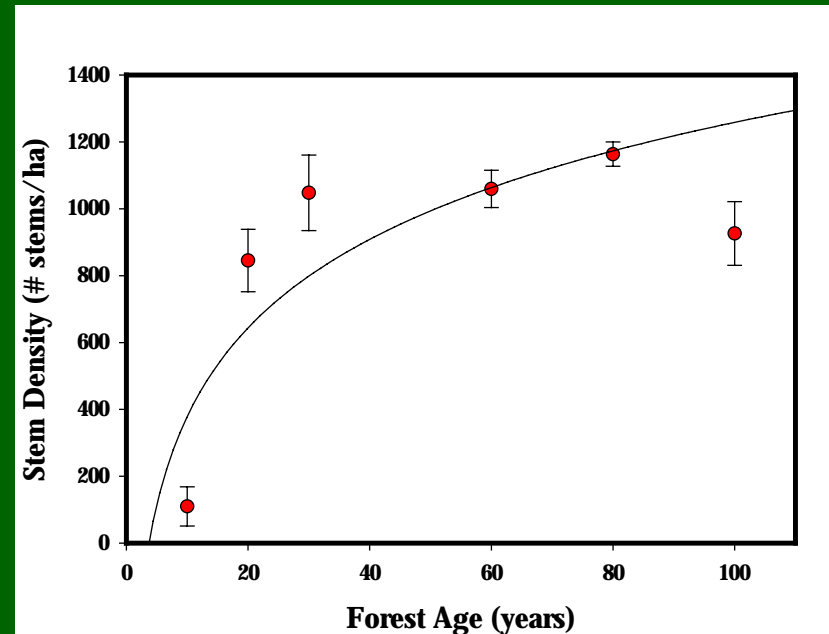
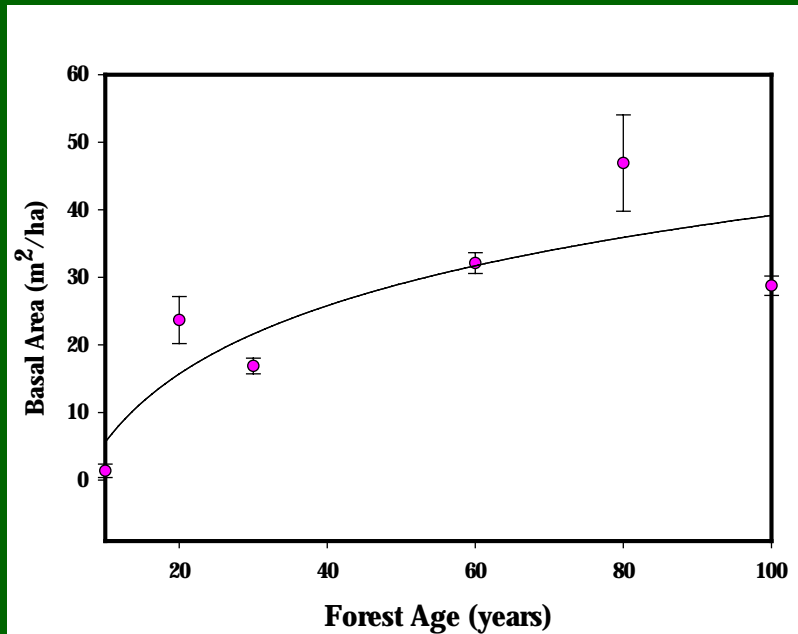


3 pits per site, n= 3 sites

Pastures did not have greater fine root biomass than forests at any depth.

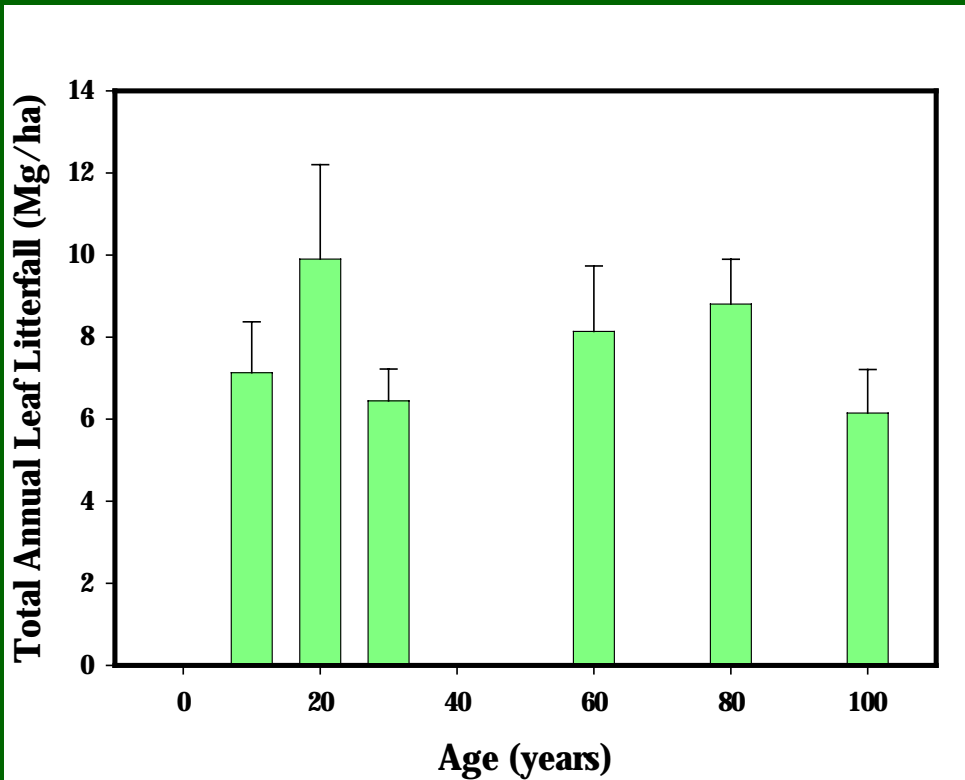


Basal Area (m^2/ha) and stem density of trees with DBH >10 cm increased with forest age.



At least 27 genera in 23 families (DBH >10 cm)

Total annual leaf litterfall did not vary with forest age.



Litter collected biweekly from 5 baskets per site.

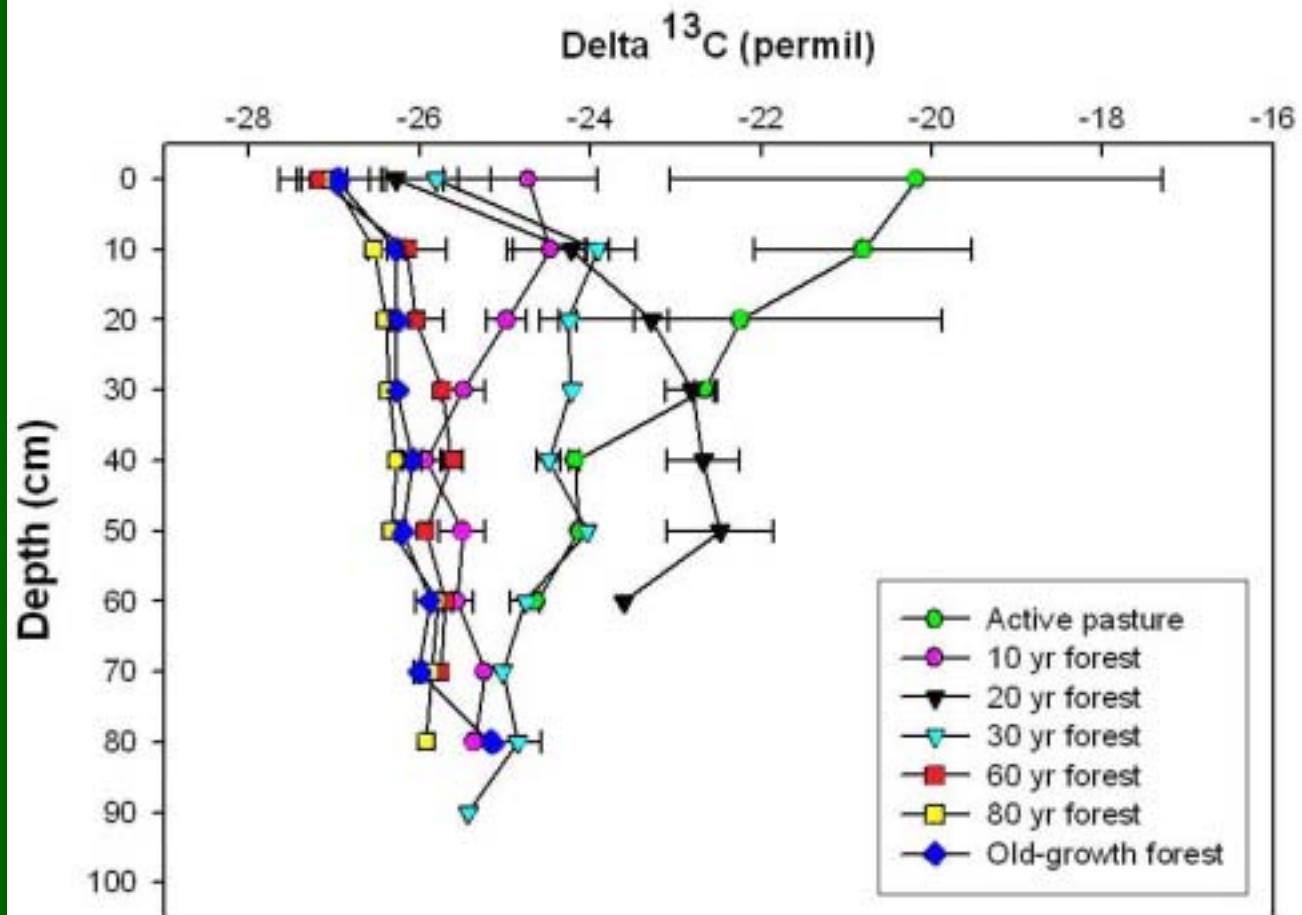
Using C Isotopes to Estimate SOM Turnover Times

- Simple mixing model to determine proportion of C4 vs. C3 derived C in SOM pool:

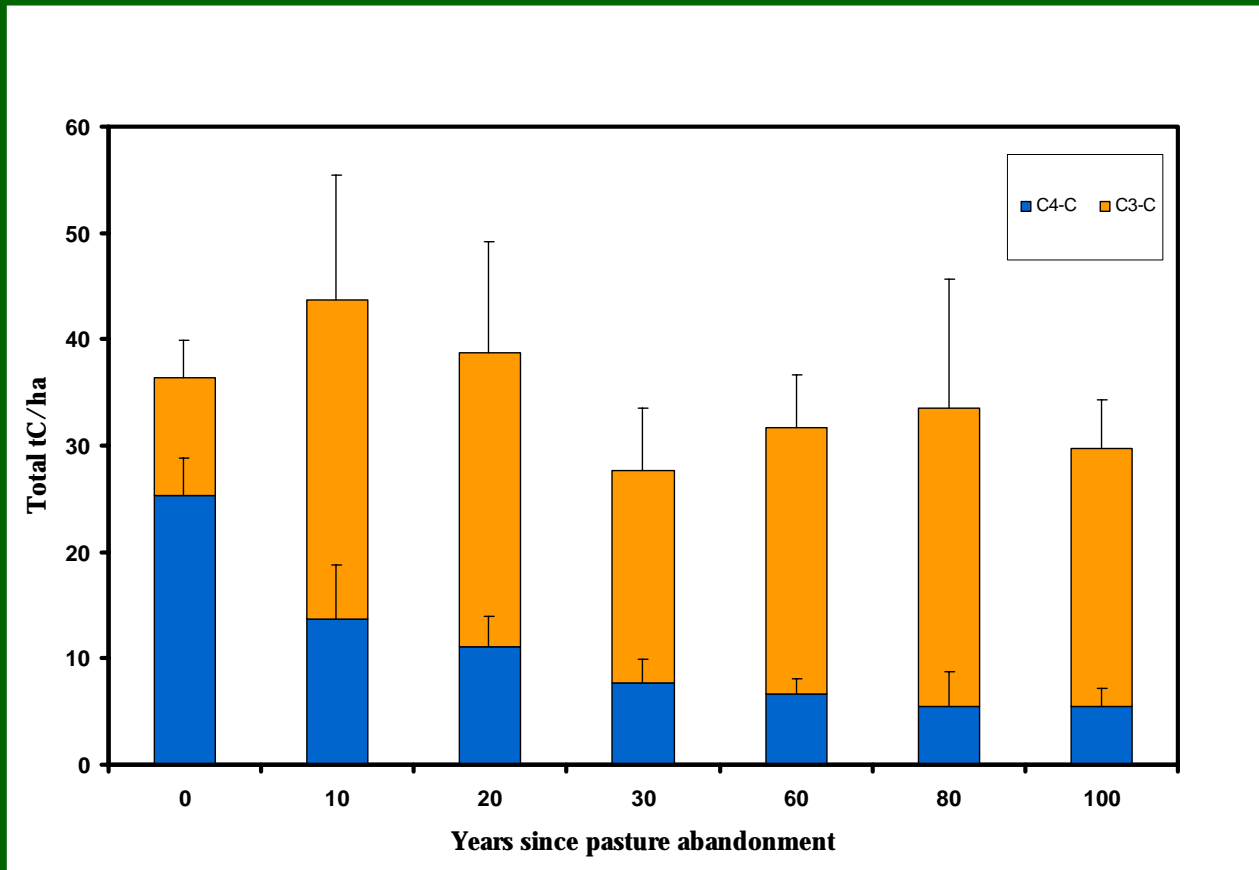
$$\begin{aligned}\%C4 &= (\delta - \delta_L / \delta_G - \delta_L) \times 100 \\ \%C3 &= 100 - \%C4\end{aligned}$$

- where δ is the $\delta^{13}C$ of the soil sample in question, δ_L is the $\delta^{13}C$ of a composite sample of forest floor and roots (or C₃), and δ_G is a composite sample of pasture grass tissues (C₄).
- “Bomb” Radiocarbon (¹⁴C) will be analyzed at CAMS-LLNL

Soil $\delta^{13}\text{C}$ -C (‰) with Depth



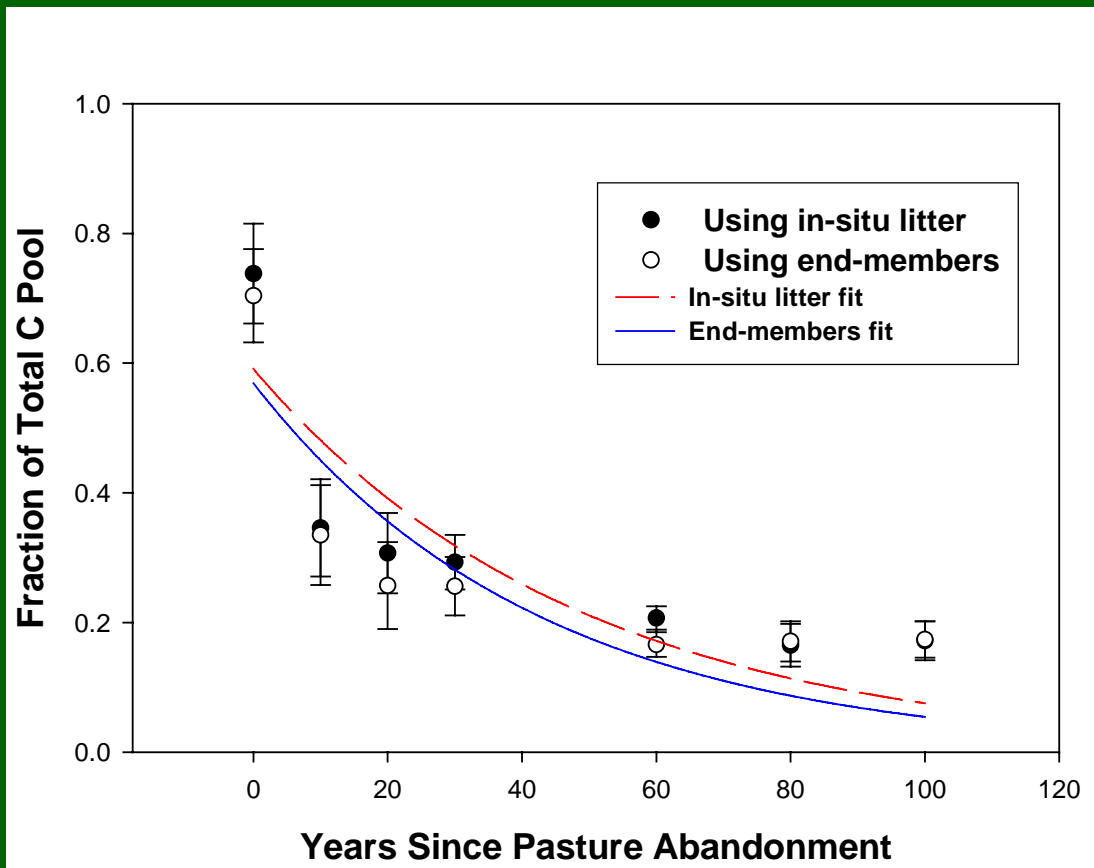
Contribution of C4-C and C3-C (in t C/ha) to total soil C pool in top 10 cm



Contribution of C₄-C to total soil C pool (0- 10 cm)

Fraction C₄-C in-situ = $0.59 - 0.021 (\text{age})$ ($r^2 = 0.74$, $p = 0.01$)

Fraction C₄-C end-members = $0.57 - 0.024 (\text{age})$ ($r^2 = 0.70$, $p = 0.02$)



Average C₄-C value of -15.02 ‰ and C₃-C value of -29.52 ‰

C Fractionation

- Experimental and modeling studies suggest that the total C pool is composed of different components, or “fractions”, with different residence times in the soil
- Attempts to separate total C pool into “fractions”, ie. stages of decomposition
- Common methods: particle size, density, aggregate-size, solubility, isotopes
- Operationally defined

Density Fractionation

- Physical separation of soil organic matter:
 - **Free Light Fraction** (f LF): identifiable leaf, root fragments, unattached organic debris (youngest)
 - **Occluded Light Fraction** (o LF): OM released after disruption of aggregates (older)
 - **Heavy Fraction** (HF): mineral associated OM (oldest)

Objective 2 : Mechanisms of soil C storage

- Examine effect of changes in soil physical structure and plant litter chemistry on the formation of stable SOM.

Hypotheses:

1.) *The primary mechanism for soil C storage during reforestation will be the development of an **aggregate hierarchy**.*

2.) *The **hydrophobic** content of plant litter will be more important than traditional measures of litter quality in the formation of stable soil C.*

H2: The hydrophobic content of plant litter will be more important than traditional measures of litter quality in the formation of stable soil C.

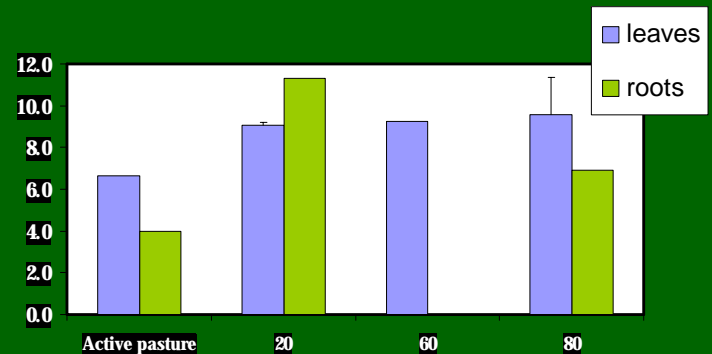
- Litter C:N, lignin:N and lignin content as measure of decomposability
- But lignin degraded in soils
- Evidence of accumulation of nonpolar C in older soil C fractions
- Recent attention to plant and soil lipids as precursors to most stable SOM

- Soil lipids: plant and microbially derived
- Plant lipids: secondary compounds, waxes, suberin, terpenoids.
- Theories of plant herbivory suggest production of these secondary plant compounds increases with forest succession

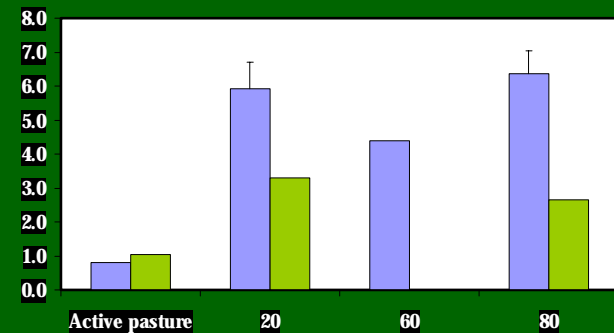
Preliminary Litter Chemistry Results

- On average, forest tissues had higher hydrophobic (NPE) and tannin concentrations than pasture tissues ($p < 0.05$).
- Leaves had higher NPE than roots across land use ($p < 0.05$).
- Aboveground pasture tissues had significantly lower levels of NPE than the 20 year old forest leaves ($p < 0.05$).

Nonpolar extractables (% dw)

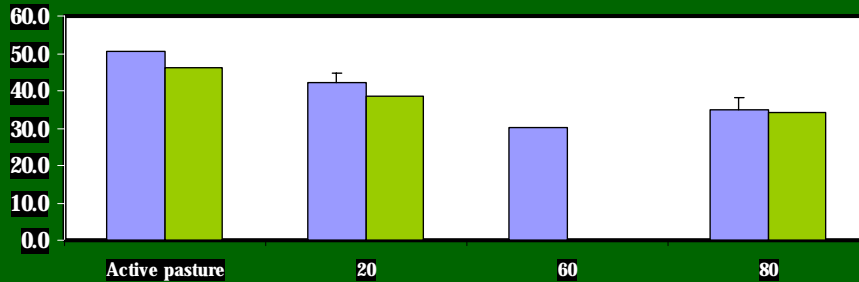


Tannins (% dw)

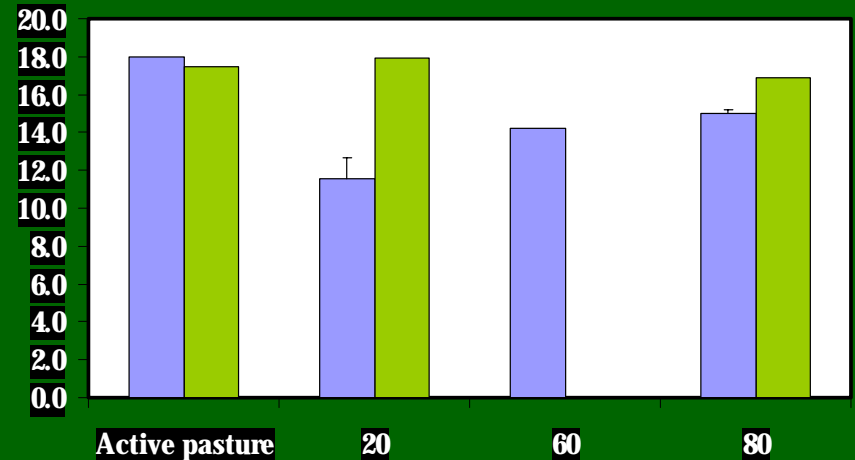


- There were no patterns in C:N and lignin:N across land use or over time.

C:N

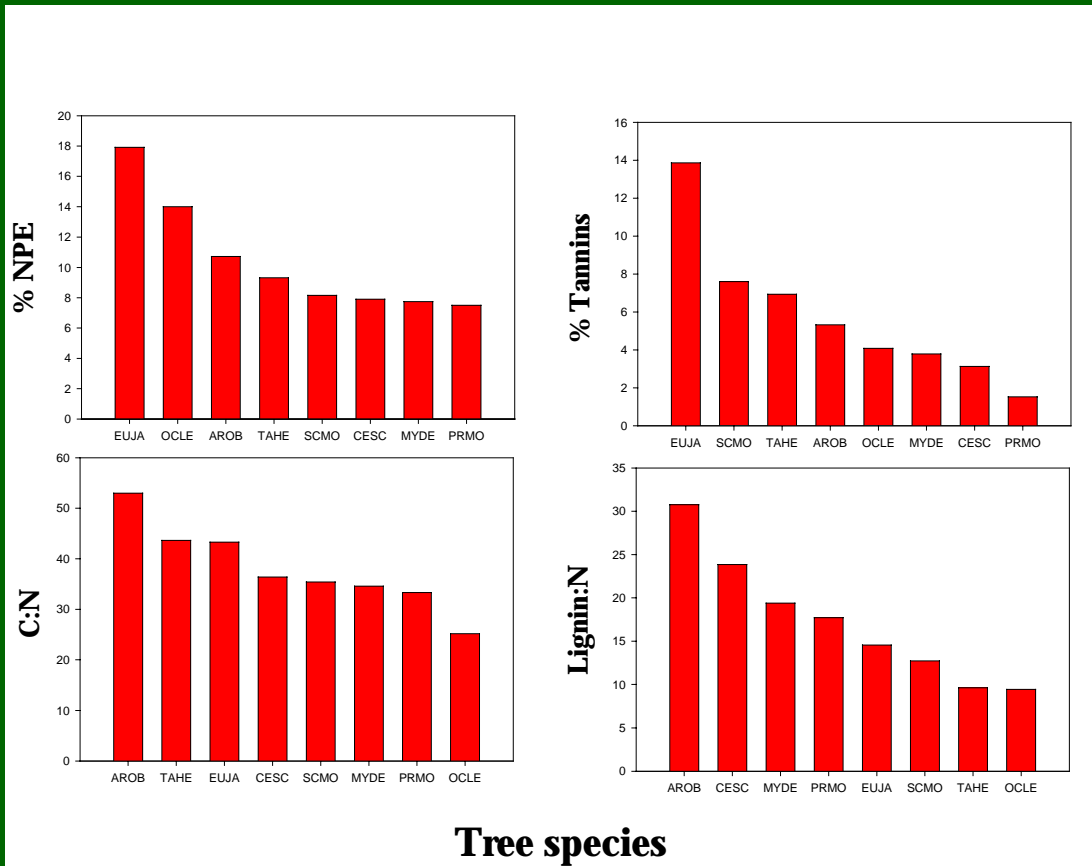


Lignin:N



Leaves (blue) and roots
(green) (0-10 cm)

Species differences in leaf tissue chemistry



Syzigium jambos (EUJA), a common invader in secondary forests, had the highest tannin and NPE concentrations; while *Prestoea montana* (PRMO), more important in old-growth forests had the lowest values for tannin and NPE.

Litter quality variables for grab samples of leaves collected at the same site from common tree species.

Species codes: SCMO, *Schefflera morototoni*; PRMO, *Prestoea montana*; TAHE, *Tabebuia heterophylla*; EUJA, *Syzigium jambos*; OCLE, *Ocotea leucoxyton*; MYDE, *Myrcia*

deflexa; CESC, *Cecropia schreberiana*.

Hydrophobic C: protected by chemical recalcitrance or physical protection? Interaction?

- Are the C fractions with slowest turnover rates hydrophobic?
- Approach
 1. Characterize and quantify hydrophobicity of SOM and litter inputs: nonpolar organic extractions, lipid analysis (GC-MS and ^{13}C -NMR)
 2. Characterize nonpolarity C in light fraction (*using XAD-8 resins*) and sorbed to minerals (^{13}C -NMR); correlate with turnover rates of fractions

3. Test for correlations between chemistry plant inputs and SOM pools, SOM turnover rates, litter decomposition rates
4. How does chemical composition of SOM/DOM affect physical protection?
 - Quantify sorptive capacity of soils at my sites
 - Perform adsorption experiments with “native” and “transplant” DOM and SOM and litter extracts

Summary

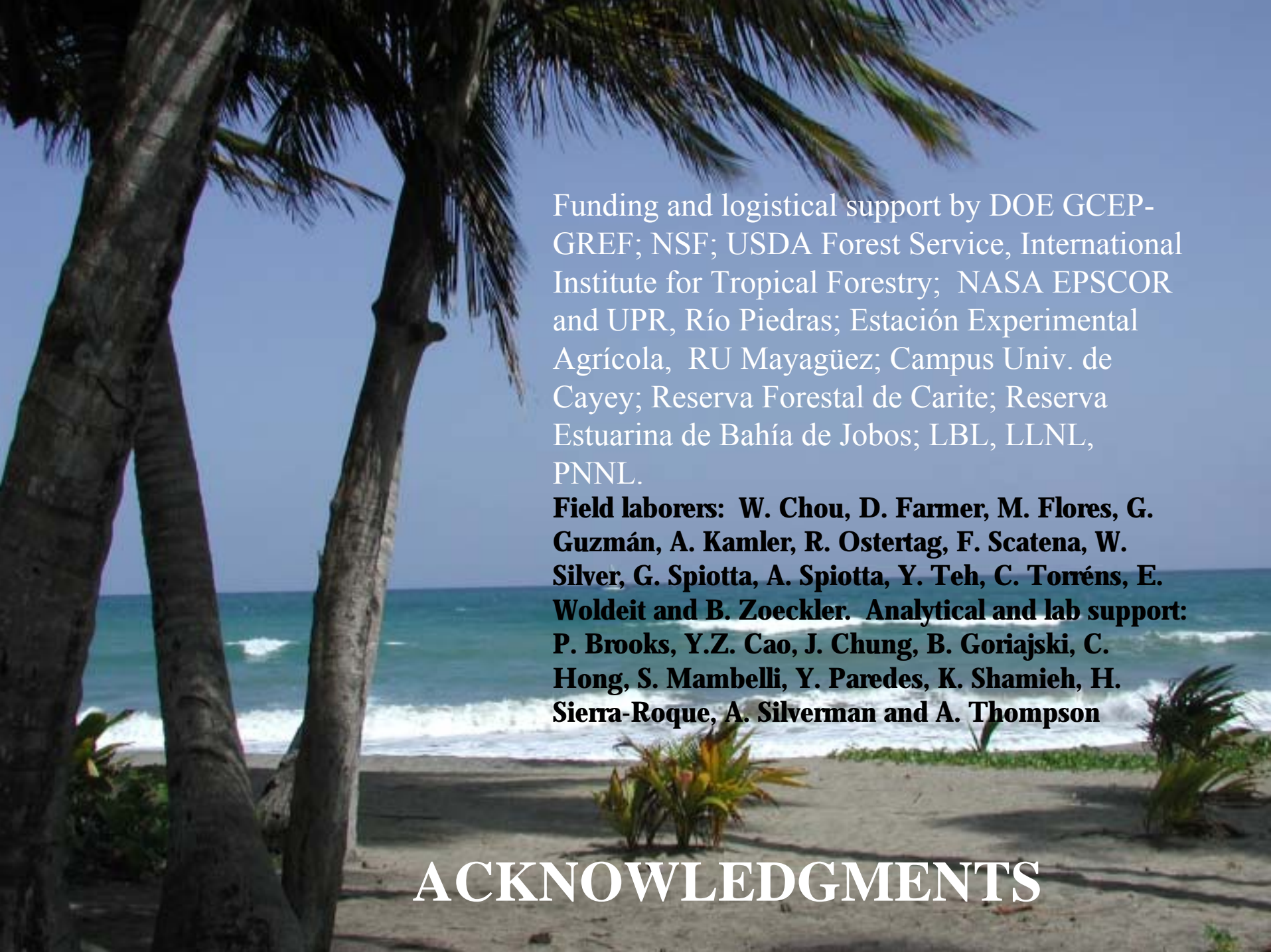
- Patterns in soil C gain and loss over time using ^{13}C and ^{14}C
- Correlations between litter quality (hydrophobicity) and soil turnover rates



Collaborators

- **Dr. Whendee Silver** (U.C. Berkeley): soil respiration and other trace gas production
- **Dr. Rebecca Ostertag** (U. of Hawaii): foliar and root litter decomposition experiments (litter vs. site quality transplant)
- **Dr. Margaret Torn** (LBNL & GREF mentor): ^{13}C - CO_2 soil respiration and ^{14}C modeling
- **Dr. Chris Swanston** (CAMS, LLNL): density fractionation and ^{14}C analyses
- **Dr. Sarah Burton** (EMSL, PNNL): ^{13}C -NMR facility and data interpretation
- **Mario Flores**: textural analyses and particle-size fractionation





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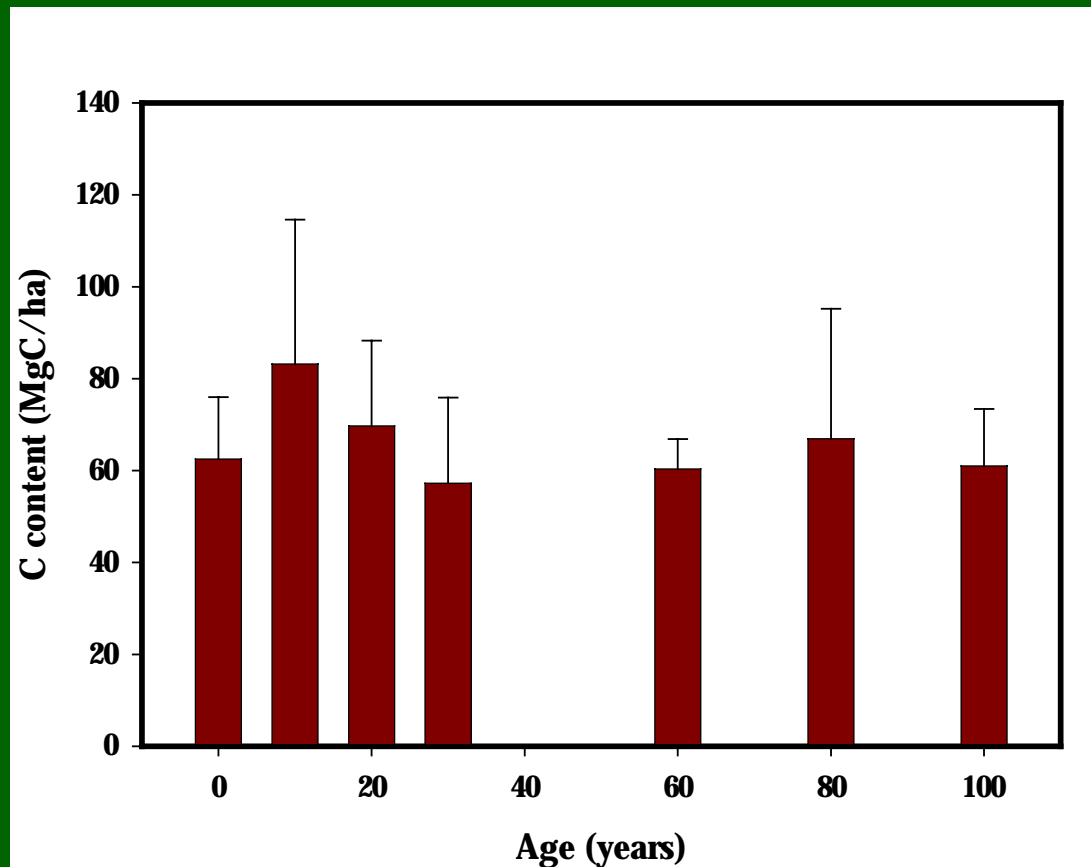
Field laborers: W. Chou, D. Farmer, M. Flores, G. Guzmán, A. Kamler, R. Ostertag, F. Scatena, W. Silver, G. Spiotta, A. Spiotta, Y. Teh, C. Torréns, E. Woldeit and B. Zoeckler. Analytical and lab support: P. Brooks, Y.Z. Cao, J. Chung, B. Goriajski, C. Hong, S. Mambelli, Y. Paredes, K. Shamieh, H. Sierra-Roque, A. Silverman and A. Thompson

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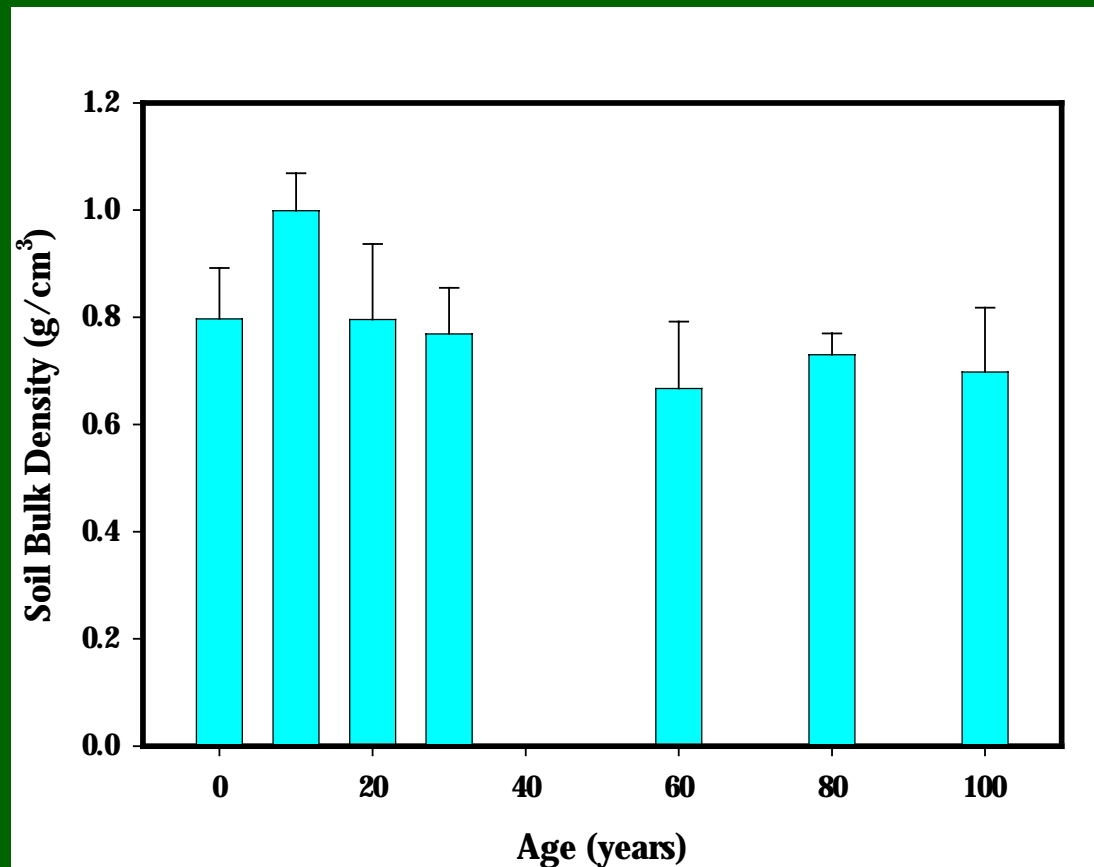
The Plant Ecologists' Café



Pastures did not differ from forests in C content in top 30 cm of soil profile.



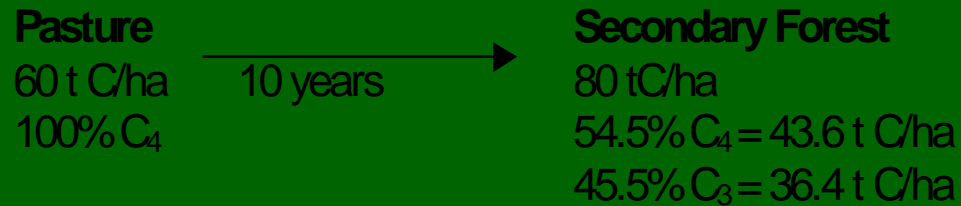
Bulk density in the 0-10 cm depth did not differ significantly. Two of the three old-growth forest sites are visited by cattle; the third site with no evidence of cattle had the lowest bulk density value recorded: 0.49 g/cm³.



Assuming a δ_L of -17‰ and a δ_G of -26‰, at our 20 year old sites:

Soil depth (cm)	%C ₃	%C ₄	C ₃ /C ₄
0-10	100	0	
10-20	76	24	3.10
20-30	67	33	2.06
30-40	74	26	2.79
40-50	55	45	1.21
50-60	50	50	1.01

Table 1. Example of proportion of C₃ and C₄ derived C for 20 year old sites.

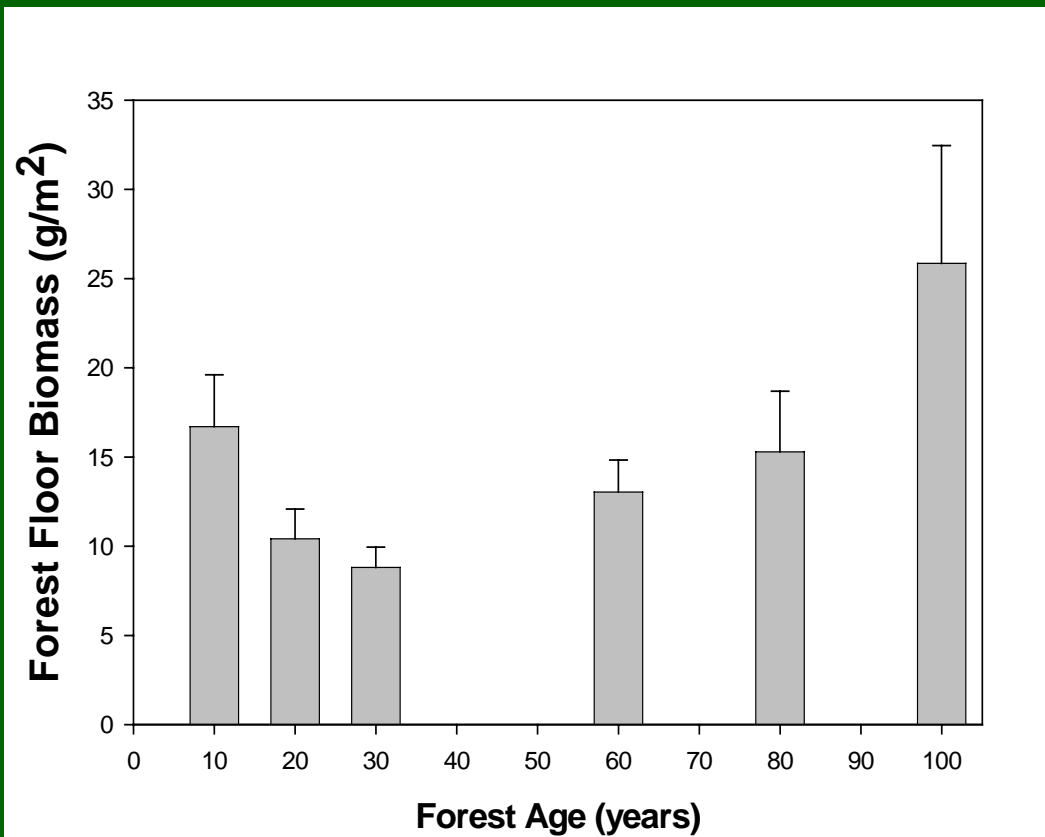


- After 10 years forest regrowth, 20 tC/ha gained, but 16.4 t C/ha of pasture-derived C was lost.

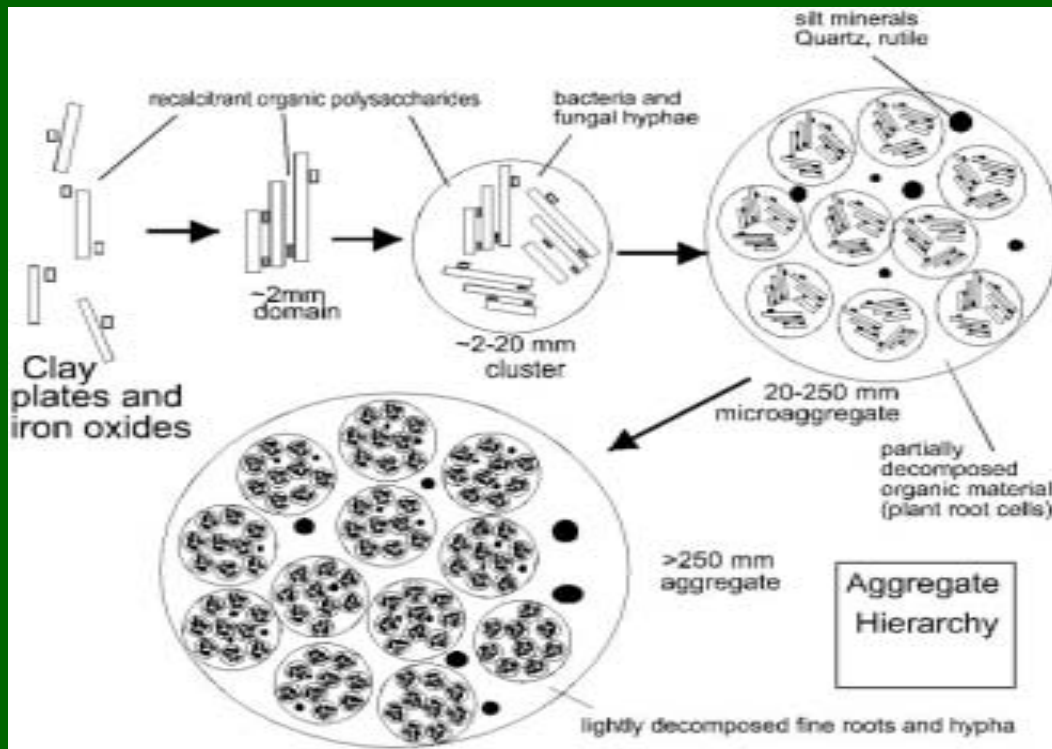
- Challenges using ^{13}C method:
 - unable to distinguish between residual “primary” forest C and new secondary forest C (both C_3)
 - uncertainties in $\delta^{13}\text{C}$ of end points, ages, turnover rates, land use history
 - assumptions inherent in “chronosequence” studies
 - simple mixing model (will try to improve)

I will also use ^{14}C and bomb carbon models to “date” soil C fractions and resolve uncertainties in turnover rates.

Average forest floor biomass in secondary forests.



H1: *The primary mechanism for soil C storage during reforestation will be the development of an aggregate hierarchy.*



- **AGGREGATE HIERARCHY:** model hierarchical contribution SOM to soil aggregate stabilization (Oades and Waters 1991)

- C protected within microaggregates where accessibility to microbes is limited or anaerobic conditions may occur
- Lower C contents in cultivated soils attributed to disruption of soil aggregates
- C within aggregates is older than C on aggregate surfaces
- CO₂ lost from disturbed aggregates

- Aggregate hierarchy thought not to be important in highly weathered tropical soils
- But recent evidence AH in Oxisols
- Recovery of aggregation post disturbance?
- Effect cattle vs pasture grasses - bulk density vs roots



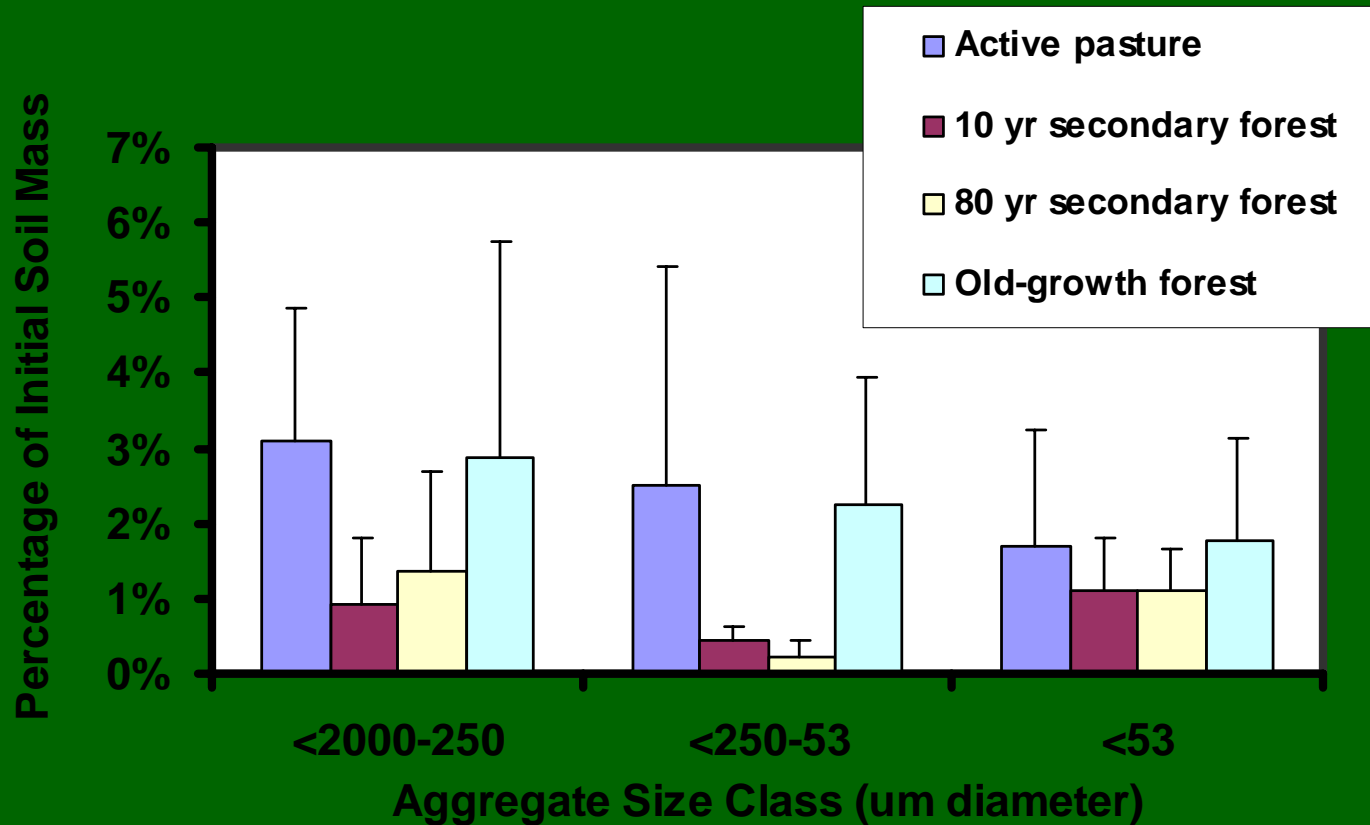
- Approach:
- Test for differences in water-stable aggregate size distribution across sites
- Test for presence of aggregate hierarchy:
 - expect total C and N to increase with size
 - expect C:N ratio to decrease from larger to smaller



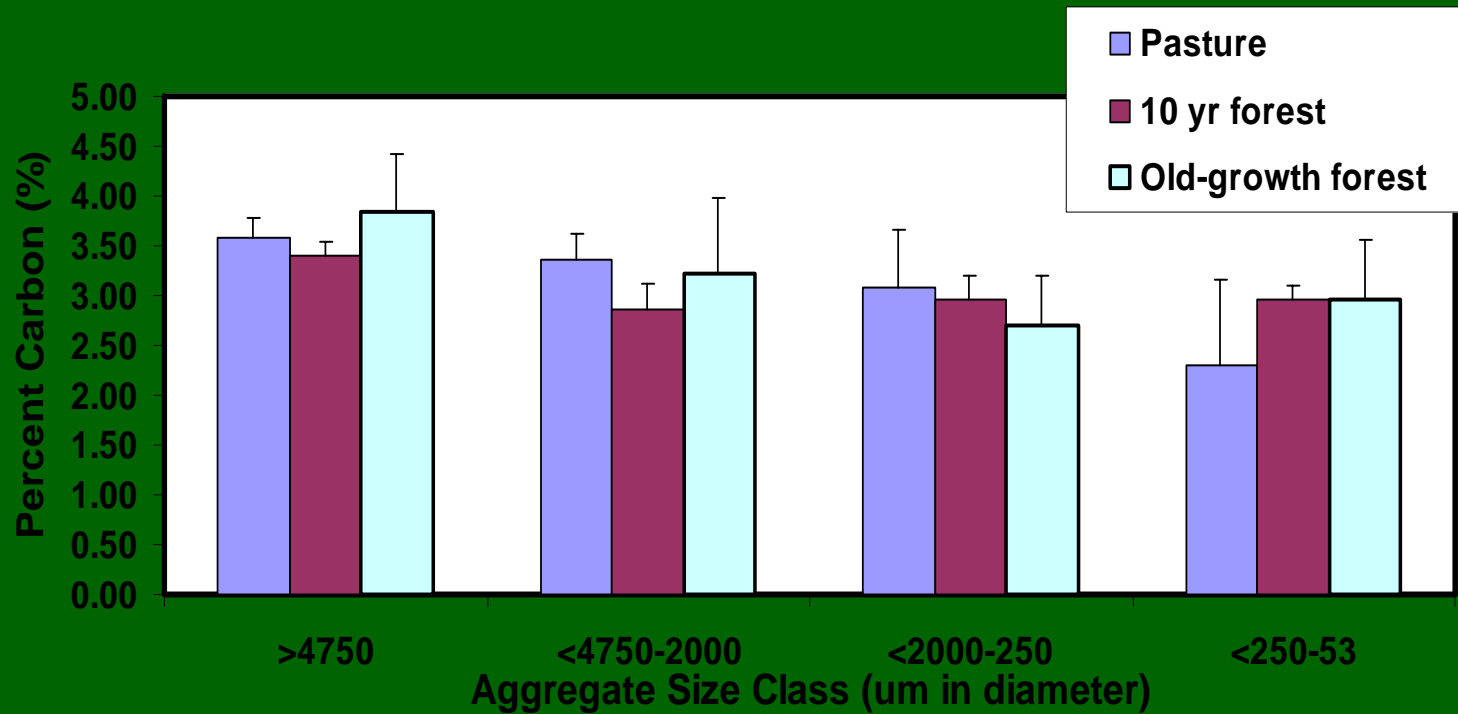
Preliminary Results:

Large macroaggregates (>4.75 mm diameter) are very stable to slaking in water for 5 min (standard method in temperate agricultural soils) so treatment did not result in differences in agg distribution and stability across sites.

Percentage of the initial mass of soil aggs > 4750 um that disassociated into smaller agg sizes and into primary particles.



C composition of different aggregate sizes



- The defining characteristics of AH are:
 - (1) a gradual breakdown of macroaggregates into microaggregates with increasing dispersing energy;
 - (2) an increase in C content with increasing aggregate size; and
 - (3) decrease in C turnover rates from macroaggregates to microaggregates (Six et al. 2000).